

Discoidin domain receptor tyrosine kinases: new players in cancer progression

Rajeshwari R. Valiathan · Marta Marco ·
Birgit Leitinger · Celina G. Kleer · Rafael Fridman

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Abstract Almost all human cancers display dysregulated expression and/or function of one or more receptor tyrosine kinases (RTKs). The strong causative association between altered RTK function and cancer progression has been translated into novel therapeutic strategies that target these cell surface receptors in cancer. Yet, the full spectrum of RTKs that may alter the oncogenic process is not completely understood. Accumulating evidence suggests that a unique set of RTKs known as the discoidin domain receptors (DDR) play a key role in cancer progression by regulating the interactions of tumor cells with their surrounding collagen matrix. The DDRs are the only RTKs that specifically bind to and are activated by collagen. DDRs control cell and tissue homeostasis by acting as collagen sensors, transducing signals that regulate cell polarity, tissue morphogenesis, and cell differentiation. In cancer, DDRs are hijacked by

tumor cells to disrupt normal cell–matrix communication and initiate pro-migratory and pro-invasive programs. Importantly, several cancer types exhibit DDR mutations, which are thought to alter receptor function and contribute to cancer progression. Other evidence suggests that the actions of DDRs in cancer are complex, either promoting or suppressing tumor cell behavior in a DDR type/isoform specific- and context-dependent manner. Thus, there is still a considerable gap in our knowledge of DDR actions in cancer tissues. This review summarizes and discusses the current knowledge on DDR expression and function in cancer. It is hoped that this effort will encourage more research into these poorly understood but unique RTKs, which have the potential of becoming novel therapeutic targets in cancer.

Keywords Discoidin domain receptor · Tyrosine kinase · Collagen · Extracellular matrix · Signaling · Cell migration · Metastasis

R. R. Valiathan · M. Marco · R. Fridman
Department of Pathology and Proteases and Cancer Program,
Wayne State University School of Medicine and Barbara Ann
Karmanos Cancer Institute,
Detroit, MI 48201, USA

B. Leitinger
National Heart and Lung Institute, Imperial College London,
London SW7 2AZ, UK

C. G. Kleer
Department of Pathology and Comprehensive Cancer Center,
University of Michigan Medical School,
Ann Arbor, MI 48109, USA

R. Fridman (✉)
Department of Pathology, Wayne State University,
540 E. Canfield Ave.,
Detroit, MI 48201, USA
e-mail: rfridman@med.wayne.edu

1 Introduction

A fundamental characteristic of metastatic cancers is the ability of tumor cells to acquire an invasive phenotype and disseminate to other organs. The acquisition of this malignant phenotype is accompanied by disruptions in the physiological interactions of tumor cells with their immediate microenvironment represented by the surrounding extracellular matrix (ECM). During normal conditions, cell–matrix interactions support cell polarity, differentiation, and survival via specific cell surface receptors that transduce cues from the ECM and activate signaling networks that support normal cellular function. In cancer, dysregulation of ECM-induced signaling disrupts normal tissue organization and

contributes to the migratory and pro-invasive programs of cancer cells. The major ECM components known to interact with almost all cell types are collagens. There are 28 different types of collagen, which collectively constitute the most abundant proteins in vertebrates. Besides their structural role as scaffolding proteins, collagens initiate cellular signaling events by activating specific cell surface receptors. The integrin and discoidin domain receptor (DDR) families are the most widely expressed collagen receptors in vertebrates. Each activates unique signaling pathways in response to collagen and elicits distinct cellular responses. A plethora of structural, functional, and clinical studies exist on integrins and on the role they play in cancer tissues. In contrast, little is known about the expression and function of DDRs in cancer progression. Therefore, in this review, we set to summarize the current knowledge on DDRs with an emphasis on their expression and functions in cancer.

2 Genomic and protein domain organization of DDRs

2.1 Genomic organization

In the early 1990s, several groups reported the cloning of a novel class of transcripts, which turned out to encode proteins with an unusual N-terminal discoidin I-like domain and a C-terminal kinase domain that was about 45% identical to that of the neurotrophin receptor, TrkA [1–10]. These novel proteins were given distinct names: DDR, TrkE, NEP, CAK, RTK-6, Ptk3, MCK-10, CCK-2, TKT, and Tyro 10. It was later recognized that these proteins were in fact either different isoforms of the same RTK or different homologs belonging to a novel family of RTKs. Based on homology of the N-terminal domain, the DDR, TrkE, CAK, RTK-6, and MCK-10 proteins were re-named as DDR1, while the CCK-2, TKT, and Tyro 10 proteins were re-named as DDR2 [11, 12]. The *DDR1* gene maps to human chromosome 6 (6p21.3) and is composed of 17 exons that are alternatively spliced to generate five different transcripts, which give rise to five distinct DDR1 isoforms. The *DDR2* gene, which maps to human chromosome 1 (1q23.3), is made up of 19 exons and encodes a single transcript, with one protein isoform.

2.2 Domain organization

DDRs are type I transmembrane RTKs that display an overall structural organization that is similar to many members of the RTK family [13], albeit with unique features (Fig. 1). The N-terminal DDR discoidin domains are homologous to discoidin I, a secreted protein from the slime mold *Dictyostelium discoideum* (reviewed in [14]). In this organism, discoidin I functions as a lectin (a carbohydrate binding protein), playing a role in cell–cell aggregation and

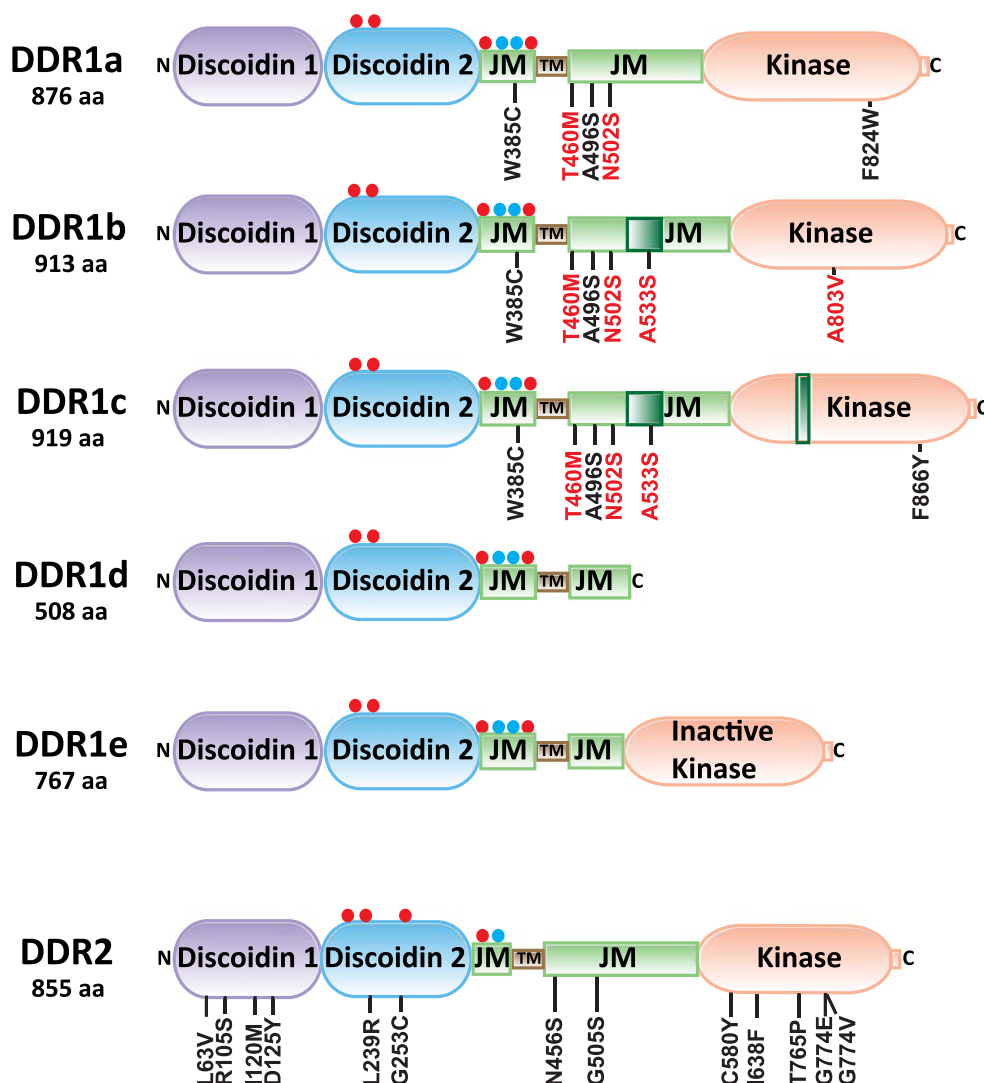
cytoskeletal organization [15]. Discoidin domains are found in several membrane and secreted proteins such as the blood coagulation factors V and VIII and the milk proteins MFG-E8 and BA46, among others (reviewed in [16]). In both DDR1 and DDR2, the N-terminal discoidin domains, referred to here as discoidin 1 (Fig. 1), have been found to bind to various types of collagens (reviewed in [17]). Besides discoidin 1, the ectodomain of DDRs contains another globular domain that is predicted to belong to the same superfamily [13], referred to here as discoidin 2 (Fig. 1). This domain is followed by an extracellular juxtamembrane (JM) region of about 50 (DDR1) or 30 (DDR2) amino acids. A single transmembrane (TM) domain comes next, followed by an unusually large cytosolic JM domain (up to 169 or 140 amino acids in DDR1 and DDR2, respectively). A catalytic kinase domain follows the cytosolic JM domain and at the very end comes a short C-terminal tail (Fig. 1). Within its intracellular region, DDR1 contains 15 tyrosine residues: seven in the JM region and eight in the kinase domain (Fig. 2). DDR2 contains 14 tyrosine residues: four in the JM region and ten in the kinase domain.

Three of the five DDR1 isoforms, DDR1a, DDR1b, and DDR1c, encode full-length, functional receptors while DDR1d and DDR1e encode truncated or kinase inactive receptors. DDR1a is generated as a result of deletion of exon 11 in the cytosolic JM domain [18]. DDR1c is generated as a result of the use of an alternate 5' splice acceptor site at the 5' intron/exon boundary of exon 14 within the kinase domain, giving rise to an additional 18 bp without open reading frame disruption [18]. The DDR1d isoform is generated through deletion of exons 11 and 12 in the cytosolic JM domain, resulting in a frame-shift mutation and a pre-maturely truncated protein [19]. In DDR1e, exons 11 and 12 are deleted, but the open reading frame is not disrupted due to an additional deletion at the beginning of exon 10 (via an alternate 5' splice acceptor site in exon 10). However, in DDR1e, the ATP binding site within the kinase domain (at the end of exon 12) is missing, rendering this isoform inactive [19].

2.3 Posttranslational modifications of DDRs

The predicted molecular mass of the full-length DDR1 isoforms ranges from ~97 to ~102 kDa. However, in immunoblots, DDR1 appears as a doublet of ~125 kDa, likely due to posttranslational modifications [20]. Similarly, DDR2 runs at an apparent molecular mass that is much higher than that predicted by its amino acid sequence. Using software that predicts glycosylation sites (<http://www.expasy.ch/tools/>), we were able to identify several potential N- and O-glycosylation sites within the DDR1 and DDR2 discoidin 2 domains (Fig. 1). Whether those sites are in fact glycosylated *in vivo* and how they affect DDR activation remains to be determined.

Fig. 1 Domain structure of DDRs. Residues that are added as a result of alternative splicing are indicated by *dark green boxes* within the corresponding domain. *Red* and *blue circles* indicate putative *N*-glycosylation and *O*-glycosylation sites, respectively. Mutated DDR residues, which were identified in samples of non-small cell lung carcinomas (*black*) and acute myeloid leukemia (*red*) are indicated. *aa* amino acids, *JM* juxtamembrane region, *TM* transmembrane region



RTKs are characterized by their ability to generate stable dimers, which is an essential requirement for receptor auto-phosphorylation [13]. While the majority of RTKs are single chain receptors that generate dimeric complexes upon ligand binding, some RTKs exist as preformed dimers. Ligand-induced RTK dimerization occurs via multiple interactions at distinct domains inducing conformational changes in the dimer that support phosphorylation of tyrosine residues in the kinase and JM domains. Studies by Curat et al. found the discoidin 1 domain to be required for dimerization of DDR1 [20]. Interestingly, Agarwal et al. and Leitinger later showed that dimerization of the DDR ectodomains is a prerequisite for ligand/collagen binding in both DDR1 and DDR2 [21–23]. Furthermore, ligand-independent DDR dimers were found to originate within the biosynthetic pathway and to be stable at the cell surface even in the absence of collagen [24–26]. Therefore, unlike the paradigm of the majority of RTKs whereby ligand binding precedes receptor dimerization, DDRs appear to exist in dimeric complexes prior to interaction with the ligand. Other studies found that

neither the extracellular nor the intracellular regions of DDR1 are required for receptor dimerization. Instead, these studies proposed that dimerization occurs via the TM region of DDR1, which was found to have a strong potential for self-association via the action of a leucine zipper motif [24]. Nonetheless, the structural and cellular mechanisms regulating DDR dimerization in the absence or presence of collagen are practically unknown.

3 Collagen binding of DDRs

3.1 Collagen specificity

DDRs are unique among RTKs because they are activated by an extracellular matrix protein, collagen (typical RTKs use soluble, peptide-like growth factors as their ligands). The DDRs only bind collagen in its native, triple-helical conformation and do not recognize heat-denatured collagen (gelatin) [11, 23]. However, triple-helical peptides containing

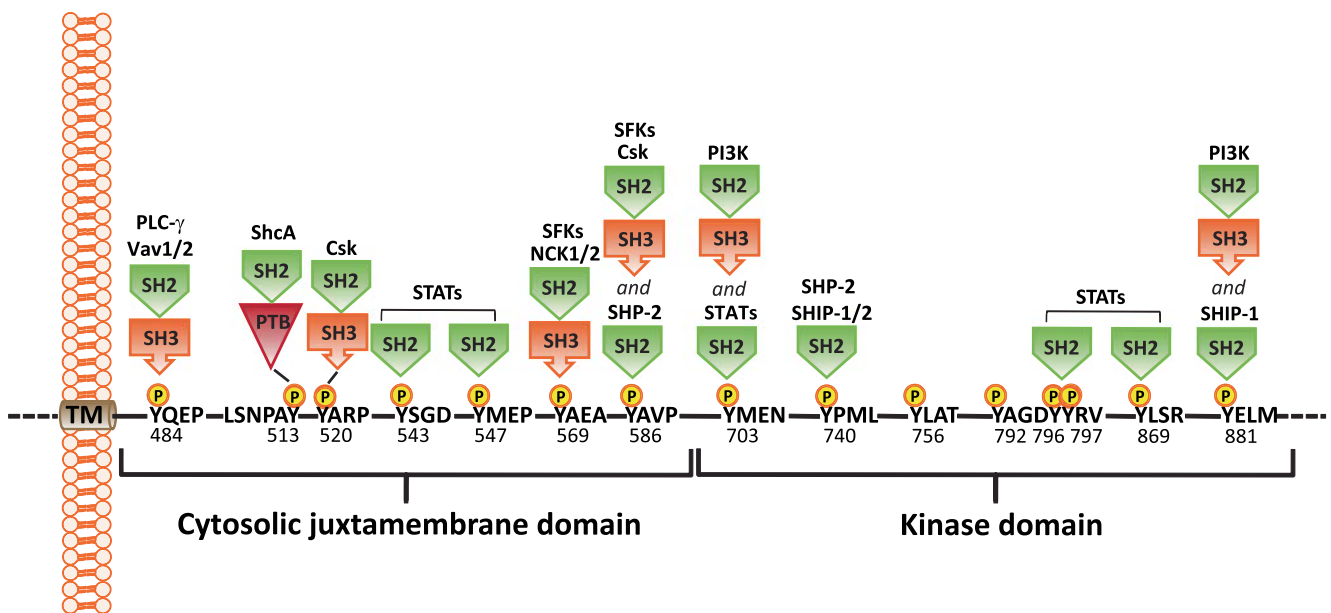


Fig. 2 Identified phospho-DDR1 interactions. The PTB domain in ShcA and the SH2 domain in Csk and Nck2 directly interact with phospho-DDR1. In the case of PLC- γ , Vav1/2, SFKs, and PI3K, which contain both SH2 and SH3 domains, the interacting domain(s) is unknown. *TM* transmembrane domain, *SH2* Src homology 2 domain, *SH3* Src homology 3 domain, *PTB* phosphotyrosine-binding domain,

PLC γ phospholipase C γ , *ShcA* SH2 containing transforming protein A, *Csk* C-terminal Src kinase, *SFKs* Src family tyrosine kinases, *PI3K* phosphoinositide 3 kinase, *SHP-2* SH2 containing protein tyrosine phosphatase 2, *SHIP-1/2* SH2-containing inositol polyphosphate 5-phosphatase 1/2, *STATs* signal transducer and activator of transcription. The indicated amino acid annotations refer to the DDR1b isoform

the collagen-binding motif of DDRs (described below) are able to induce receptor activation, indicating that the supra-molecular structure of collagen is not required for DDR signaling [27]. Both DDRs display broad collagen specificity and are activated by many different collagen types, with fibrillar collagens acting as ligands for both receptors [11, 12]. The DDRs have distinct preferences for certain types of collagens. DDR1, but not DDR2, binds to the basement membrane collagen IV [11, 12], while DDR2 seems to preferentially bind collagen II [28] and collagen X [29]. DDR1 can also bind collagen VIII [30], but it is not known whether DDR2 shares this property. Similar to collagen-binding integrins, the DDRs recognize specific amino acid motifs in collagen. Initial mapping of DDR2 binding sites was performed with recombinant triple-helical variants of collagen II, which mapped a specific DDR2 binding site to the second quarter of its collagenous (COL) domain [28]. More detailed studies, utilizing libraries of triple-helical peptides, the so-called Collagen Toolkits [31], uncovered a six amino acid motif, GVMGFO (O is hydroxyproline), as a binding motif for both DDRs [27, 32]. The GVMGFO motif is present in the fibrillar collagens I–III. DDR2 has additional binding sites in collagens II and III [27, 32], but their exact sequences have not yet been determined. The distinct preferences of DDRs for various collagen types suggest that migrating cancer cells may deploy different DDRs during invasive processes through different collagen matrices.

3.2 Collagen-binding sites for DDRs

The DDR collagen-binding sites are entirely contained within their discoidin 1 domains, as shown by *in vitro* collagen-binding experiments using recombinant extracellular domains [23]. The DDR discoidin 1 domains show a high degree of conservation with 59% sequence identity. Initial mutagenesis experiments mapped the collagen-binding sites to three spatially adjacent surface-exposed loops that are highly conserved between the DDRs [23, 33]. The structure of the un-liganded DDR2 discoidin 1 domain was subsequently determined by NMR, and the collagen-binding site was identified by transferred cross-saturation experiments and mutagenesis [34]. The collagen-binding site is in the form of a trench created by five protruding loops at the “top” of the discoidin 1 domain, opposite the disulfide-linked chain termini. The identification of the GVMGFO motif as a DDR ligand enabled the crystal structure determination of a complex between the DDR2 discoidin 1 domain and a triple-helical peptide encompassing the GVMGFO motif [35]. This structure revealed the apolar GVMGFO motif to be accommodated in an amphiphilic binding pocket. The main collagen-binding residues defined by this structure are strictly conserved in DDR1, which is consistent with both receptor types binding to fibrillar collagens. Several DDR2 residues at the periphery of the GVMGFO peptide-binding interface are not conserved in DDR1. These residues are responsible for the distinctive collagen-binding specificity

of the DDRs. Replacing these amino acids in DDR2 with the corresponding DDR1 residues created a DDR2 construct that was able to bind collagen IV [32].

4 DDR activation and signaling

4.1 Mechanisms of DDR activation/phosphorylation

Like all RTKs, the DDRs undergo receptor autophosphorylation upon ligand binding, but in the DDRs, contrary to most RTKs, this process is unusually slow and sustained [11, 12]. However, the structural, biochemical and cellular mechanisms behind the slow rate of DDR activation remain unknown. Recently, Mihai et al. proposed a model of receptor activation, which may partly explain the slow and sustained phosphorylation of DDR1. Using CFP and YFP-labeled DDR1 proteins, Mihai et al. showed that collagen exposure induces rapid receptor aggregation, which is then followed by internalization of receptor into early endosome vesicles [25]. The internalized DDR1 then recycles back to the cell surface with a time frame suggestive of intracellular phosphorylation. Although this model provides an explanation for the unusually slow kinetics of DDR1 activation, the exact composition of the ligand-induced, higher-order, DDR1 oligomers are yet to be defined. Moreover, the mechanisms that trigger DDR1 endocytosis and the nature of the endocytic pathways remain unknown.

How collagen binding to the DDR discoidin 1 domain leads to the activation of the cytosolic kinase domain is not clear. It is assumed that collagen-induced transmembrane signaling involves substantial conformational changes within the dimer. Structural studies showed that DDR1 activation requires a leucine zipper motif in the TM domain [24], and it is possible that the conformational change upon collagen binding could involve a rotation of the TM helices within the dimer. However, further structural studies will be necessary to understand the nature of these conformational changes. Other open questions are whether TM signaling requires one or both discoidin 1 domains within the preformed dimer to be occupied by ligand and how auto-inhibition of the inactive dimer is achieved. Although the observation that DDRs exist in preformed dimers is consistent with a model of auto-inhibition of phosphorylation, which can only be overcome by ligand binding, studies using the phosphatase inhibitor pervanadate showed a significant and rapid phosphorylation of DDR1 in the absence of added ligand [36, 37]. These findings suggest a rapid turnover of DDR1 between phosphorylated and unphosphorylated states. Whether the observed rapid phosphorylation induced by the presence of pervanadate is due to the release of a phosphatase-mediated inhibition mechanism is not known. Collagen-evoked DDR1 activation is also

stimulated by interacting signaling pathways. For instance, the Wnt5a/Frzld pathway regulated by TGF β was shown to be necessary for maximal DDR1 phosphorylation in breast epithelial cells [38–40]. Additionally, Src has been shown to be required for DDR1 phosphorylation in breast epithelial and vascular smooth muscle (VSM) cells [39, 41]. How these signaling molecules contribute to DDR1 activation is still unclear. In the case of DDR2, Src has been shown to contribute to receptor activation in COS7 cells and in activated rat hepatic stellate (HS) cells, DDR2 was shown to form a complex with Src [42]. *In vitro*, Src phosphorylates DDR2 on three tyrosine residues, Y736, Y740, Y741, within the DDR2 kinase domain activation loop, which results in intramolecular autophosphorylation of DDR2 at other tyrosine residues [43]. Substitution of Y740 (within the activation loop of the DDR2) for phenylalanine results in constitutive DDR2 kinase activity suggesting that this residue plays a role in autoinhibition of DDR2 phosphorylation [43]. Thus, DDR2-Src interactions may play a key role in DDR2-initiated signaling.

4.2 DDR binding partners

Upon collagen binding, DDRs undergo autophosphorylation at multiple tyrosine residues within the cytosolic JM and kinase domains. Phosphorylation of tyrosine residues in a precise order recruits cytoplasmic signaling molecules containing Src homology-2 (SH2) and phosphotyrosine-binding (PTB) domains, which in turn assemble protein complexes that serve to transduce receptor signals. In DDR1b and DDR1c, the cytosolic JM region and the kinase domain possess 15 tyrosine residues that can potentially undergo phosphorylation and serve as docking sites for SH2/3 or PTB containing adaptor proteins (described in Fig. 2). Several adaptor proteins have been shown to bind DDR1 in response to collagen stimulation, and in some instances, the phosphotyrosine residues involved in binding were also identified (Fig. 2). For example, ShcA [11], the p85 α subunit of PI3K [37, 39], Nck1/2 [44], SHP-2 [44, 45], C-terminal Src kinase (Csk) [46], non-muscle myosin heavy chain (NMHC)-IIA [47], and the focal adhesion kinase (FAK) homolog, Pyk2 [48], were all found to associate with activated DDR1. Most of these interactions were confirmed in a recent proteomic study, which investigated the phosphotyrosine interactome of DDR1 in pervanadate treated SF268 human glioblastoma cells overexpressing DDR1b and in human placenta using immobilized DDR1 phosphotyrosine peptides [36]. This study also showed that Stat1a/b, Stat3, and Stat5 directly bind to various phosphotyrosine residues in DDR1, suggesting that activated DDR1 possibly brings SHP-2 and its substrates in close proximity. Other DDR1 binding proteins identified by this approach include RasGAP, the guanine nucleotide exchange factors Vav2 and Vav3, the adaptor protein CRKII,

and the phosphatase SHIP2 [36]. Several DDR1-interacting partners that bind DDR1 regardless of its phosphorylation status, have also been identified. These include DARPP32 [49], KIBRA [50], Syk [51], Notch1 [52], E-cadherin [53, 54], and the Par3/Par6 cell polarity proteins [54]. In the case of DDR2, ShcA was reported to bind at pY471 following collagen I stimulation [43], and indirect evidence suggests that DDR2 may constitutively interact with Src [42], which may also be phosphorylated by DDR2 [55].

4.3 DDR-initiated signaling

Figure 3 summarizes the downstream signaling events regulated by DDRs, with emphasis on those identified in cancer cells. For DDR signaling in non-malignant cells, the reader is encouraged to see the following studies [45, 55–62]. In human breast and colon carcinoma cell lines, DDR1 activation triggers pro-survival Ras/Raf/ERK and PI3K/Akt pathways resulting in upregulation of anti-apoptotic Bcl-xL and survival under conditions of genotoxic stress [63]. DDR1

activation also increases NFkB DNA binding activity and cyclooxygenase-2 expression in human breast cancer cells, leading to increased chemoresistance [64]. In colon cancer cells, DDR1 forms a complex with Notch 1. DDR1 activation triggers Notch1 cleavage by γ -secretase, generating the Notch 1 intracellular domain, which translocates to the nucleus and upregulates pro-survival genes such as *Hes1* and *Hey2* [52]. In pancreatic cancer cells, DDR1 activation, in conjunction with integrin β 1, triggers a p130CAS/JNK pathway that results in upregulation of N-cadherin and epithelial to mesenchymal transition (EMT)-like cell scattering [48]. Although there is a plethora of studies on DDR2 signaling in non-malignant mesenchymal cells (for example [55, 60–62]), DDR2 signaling pathways in cancer cells are practically unknown.

4.4 DDR-integrin cross talk

As malignant cells navigate through the collagen matrix, they must adapt their response to the structural and bio-

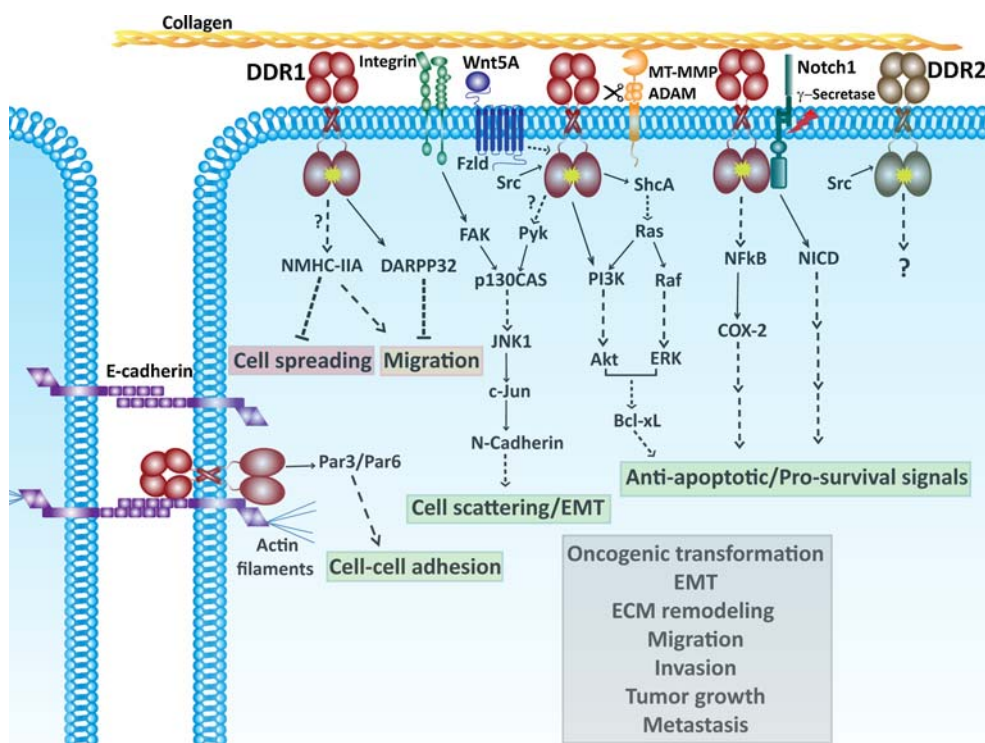


Fig. 3 Reported DDR-initiated signaling pathways in cancer cells. *Solid lines* indicate direct interactions or effects. *Dashed lines* represent indirect interactions or effects that are mediated through one or more intermediate steps. Unknown interactions (direct or indirect) are indicated with a *question mark*. *Arrows* pointing to DDRs indicate pathways involved in DDR activation and *arrows* pointing away from DDRs indicate pathways triggered by activated DDRs. Activated DDRs are indicated with a *yellow star*. The *red box* depicts processes

that are suppressed by DDR signaling. The *green box* depicts processes that are promoted by DDRs. The *orange box* depicts processes that are either suppressed or promoted by DDR signaling under different contexts. The *gray box* depicts processes that are associated with DDR expression/function, but the signaling pathways or mechanisms activated by DDRs in these processes have not been worked out yet. NICD, Notch1 intracellular domain

chemical nature of their new environment by commanding the activity of two completely different types of collagen receptors: the integrins and the DDRs. It is therefore important to know what the specific contributions of these receptors are and the potential contribution of receptor cross talk in the response of cells to the collagen matrix. In some instances, integrins and DDRs may elicit antagonistic effects, which must be integrated to accomplish the proper cellular response. Because cells generally express both integrins and DDRs, several studies have investigated whether DDR activation is coupled with integrin signaling. Although it was observed that DDR1 activation by collagen does not require the activity of collagen-binding integrins [65], some of the downstream signaling pathways activated by DDR1 appear to intersect with integrin-activated pathways. In MDCK cells, DDR1 antagonizes integrin-FAK-Cdc42-mediated cell spreading [58] and integrin-STAT1/3-mediated cell migration [45]. In pancreatic cancer cells, DDR1 and integrin $\beta 1$ signals cooperate to induce N-cadherin upregulation and cell scattering/EMT-like changes in response to collagen I [48]. In this system, DDR1 activates Pyk2 while integrin $\beta 1$ activates FAK. p130CAS, a signaling scaffold protein known to bind to FAK and Pyk2, also binds DDR1 in pancreatic cancer cells. The Pyk/FAK-p130CAS complex activates JNK1, which then upregulates N-cadherin through c-Jun, leading to cell scattering (Fig. 3) [48]. Also, collagen I, through activation of integrin $\alpha 2\beta 1$ and DDR1, plays a role in maintaining an undifferentiated state in mouse embryonic stem cells [66]. In this context, signals downstream of DDR1 and integrin cooperate to promote self-renewal via cell-cycle regulation. Specifically, integrin $\alpha 2\beta 1$ activates integrin-linked kinase, which then triggers Notch cleavage and Gli-1 upregulation. DDR1 on the other hand activates PI3K/Akt and Ras/ERK. Gli-1 and ERK are both required to activate Bmi-1, which then downregulates p16 and upregulates pRb allowing cell-cycle progression to occur in these stem cells [66]. Based on these examples, one can postulate that DDRs and integrins may play cooperating or antagonizing roles at different stages of cancer progression. Thus, it will be important to delineate the relative contributions of these two collagen receptors at various stages of cancer progression and in different cancer types in order to effectively design novel anti-tumor therapeutic regimens targeting these receptors.

5 Regulation of DDR expression

5.1 Transcriptional and posttranscriptional regulation

Transcriptional regulation of DDR1 expression is mediated in part by the Ras/Raf/ERK signaling pathway in a variety

of normal and cancer cells. In human T cells, for example, T cell receptor activation induces DDR1 expression through Ras/Raf/ERK and protein kinase C-dependent pathways [67]. In primary lung fibroblasts, collagen I upregulates DDR1 expression through a mechanism that is dependent on DDR2 activation, phospho-JAK2 recruitment to DDR2 and ERK1/2 activation, leading to increased recruitment of the polyoma enhancer A-binding protein 3 to the DDR1 promoter [68]. This suggests that the interactions of fibroblasts with collagen I require a coordinated regulation and function of both DDR receptors. DDR1 activation itself also results in Ras/Raf/ERK signaling and could therefore positively regulate its own expression, as shown in MCF7 breast cancer and HCT116 colon carcinoma cell lines [63]. Several reports have shown that induction of an EMT phenotype results in transcriptional downregulation of DDR1 in MDCK cells and in human bronchial and breast epithelial cells [69–71]. Consistent with these findings, the *DDR1* promoter contains potential binding sites for the EMT-associated transcription factor, Zeb1 [71]. However, the relative role of EMT transcription factors, including Zeb 1, in the regulation of DDR1 expression needs to be further explored.

The *DDR1* genomic sequence also contains a functional p53 regulatory element [63, 72]. Genotoxic stress induced either by ionizing radiation or chemotherapy upregulates DDR1 expression in a p53-dependent manner [63, 64, 72]. Similarly, overexpression of the DNA repair protein XRCC3 results in DDR1 upregulation by an as yet unknown mechanism [73]. A recent study in oligodendrocytes found that the *DDR1* gene contains an hnRNP A2 response element-like sequence, which may be involved in alternative splicing and nuclear export of DDR1 mRNA [74]. Also, DDR1 mRNA is a direct target of the microRNA miR-199a-5p in human hepatocellular carcinoma cells [75].

Several transcription factors/complexes have been implicated in the regulation of DDR2 expression in different systems. During osteogenic differentiation, the ATF4-C/EBP β transcription factor complex induces DDR2 upregulation [62]. In rat VSM cells, hypoxia or hyperbaric oxygen increases Myc-Max DNA binding activity in the *DDR2* promoter region thereby increasing DDR2 expression [76, 77]. In nasopharyngeal carcinomas, which are commonly associated with Epstein–Barr virus (EBV) infection, upregulation of DDR2 has been linked to direct activation of the *DDR2* promoter by the EBV-Z transactivator protein [78]. DDR2 mRNA and protein expression are upregulated in HS cells undergoing activation during liver injury [79]. Inhibition of HS cell activation by overexpression of miR-29b, which directly targets collagen I mRNA, results in the concomitant downregulation of DDR2 mRNA, suggesting a relationship between collagen I expression and DDR2

levels [80]. Acquisition of an EMT phenotype in MDCK and human breast epithelial cells has also been shown to induce DDR2 expression [69, 71]. However, the transcription factors and mechanisms involved in upregulation of DDR2 during EMT have not been investigated.

5.2 Posttranslational regulation: protease-mediated regulation

The DDRs are type I transmembrane proteins and as such are regulated by a variety of biochemical and cellular mechanisms that govern transmembrane protein trafficking including insertion into membrane microdomains (lipid rafts), apical/basal targeting, endocytosis, recycling, and degradation. At the time of this writing, there are no reports on these basic aspects of DDR regulation. There is evidence, however, on protease-mediated ectodomain shedding of DDRs, a process that can profoundly affect receptor function. A large number of membrane-anchored proteins including RTKs can be cleaved by proteases to release the extracellular domains [21, 81]. This process, known as “ectodomain shedding”, provides an extra level of regulation to membrane-anchored proteins by depleting functional ectodomains from the cell surface. In the case of RTKs, ectodomain shedding can significantly alter signaling by reducing access of the ligand to its cognate receptor. In turn, the soluble, ligand-binding ectodomain may elicit a variety of biological functions in the extracellular space and possibly influence the membrane-anchored receptor in different ways. DDR1 was shown to undergo ectodomain shedding in a constitutive [10] and collagen-induced manner [82, 83] in a process that requires metalloprotease activity. It was reported that DDR1 is cleaved within the extracellular JM region generating two phosphorylated C-terminal fragments of 58 and 62 kDa consistent with the existence of two cleavage sites [82]. Collagen-induced shedding of DDR1 in T47D breast cancer cells is reduced by a Src inhibitor suggesting a role for Src in this process [83]. The nature of the metalloprotease(s) involved in DDR1 shedding has yet to be identified. However, DDR1 cleavage is sensitive to inhibitors of MMPs and ADAMs [83], two metalloprotease families known for their ability to accomplish ectodomain shedding. Data from our laboratory implicate the transmembrane membrane type matrix metalloproteinase MT1-MMP (MMP14), MT2-MMP (MMP15), and MT3-MMP (MMP16) in the shedding of DDR1 in a process that is independent of collagen and involves cleavage within the extracellular JM region at two specific sites. Moreover, cleavage of DDR1 by these MT-MMPs blocks DDR1 phosphorylation by collagen (manuscript in preparation). So far, there are no reports on shedding of DDR2, and in our hands, DDR2 is not shed by these MT-MMPs. This suggests that DDRs are differentially regulated by membrane-anchored

collagenases, which may have implication for cell–collagen interactions during matrix remodeling. Although the functional consequences of DDR1 shedding need to be determined, this process may serve to control the pool of functional receptors at the cell surface thereby influencing collagen signaling. Alternatively, soluble DDR1 ectodomains may alter collagen fibrillogenesis and organization [22, 84, 85]. Collagenases like the MT-MMPs may also influence DDR signaling by affecting the structural integrity of the collagen matrix at areas of cell–matrix contact. Indeed, an early study showed that collagen degradation inhibits DDR1 activation [11]. Thus, the ability of metalloproteases to regulate DDR integrity and function highlights a potential DDR/protease axis, which may play a key role during intense proteolytic remodeling of the matrix by migrating cancer cells.

6 DDR expression and mutations in cancer tissues

The identification of DDRs and the realization that they are unique RTKs promoted investigation of their expression, phosphorylation, and mutational status in various cancer types, using a variety of approaches. However, several issues need to be taken into consideration when evaluating DDRs in cancer tissues. For instance, due to the limitations of the currently available DDR antibodies, immunohistochemical methods cannot provide information on receptor activation status, expression of distinct DDR1 isoforms, and/or presence of mutated DDRs. Analyses of RNA expression by real-time PCR or searches in current databases such as OncoPrint™ (Compendia Bioscience, Ann Arbor, MI, USA) provide a glimpse on the associations between DDR expression and a variety of cancer-related parameters. However, these analyses are limited because posttranslational regulation of DDRs is a key determinant of receptor function. Analyses of receptor phosphorylation status by phosphoproteomic approaches provide information on the pool of phosphorylated receptor within the sample. However, these approaches may be limited for DDR detection because DDRs exhibit very slow kinetics of phosphorylation, which may influence receptor capture. Moreover, DDR activation may not be required to support tumor cell invasion [54]. Several studies specifically addressed the mutational status of DDRs (discussed below), and in some cases, the functional consequences of these mutations were addressed. However, the downstream signaling pathways activated by these mutations and whether they are “driver” or “passenger” mutations [86] remains unclear. With these issues in mind, we summarize the results of published studies that have specifically focused on DDR expression, function and/or mutational status in different cancer types (below and Table 1).

Table 1 Summary of DDR expression in human cancer tissues

Cancer	DDR1		DDR2	
	High	Low	High	Low
Breast	Primary tumors and metastasis-containing lymph nodes ^a [95]. Invasive ductal carcinoma ^b [54, 97, 175]	Invasive lobular carcinoma ^b [97, 175]. Middle to high-grade carcinomas ^a [96]	NR	NR
Lung	Non-small cell lung carcinomas [88] ^a , [89–91] ^b			Non-small cell lung carcinomas ^a [88] NR
Brain	High-grade primary neuroepithelial and metastatic tumors ^a [99]. High-grade gliomas ^{a,b} [101]. GH- or PRL-producing pituitary adenomas and macroadenomas ^b [102]. Malignant GBMs ^{a,b} [100]	ACTH- or TSH-producing pituitary adenomas, non-functioning adenomas, and microadenomas ^b [102] Benign and borderline ovarian tumors ^a [8]. Low-grade and stage serous ovarian cancers ^{a,b} [104]	NR	NR
Gynecological	Malignant ovarian tumors (moderate, poorly, and well-differentiated) ^a [8]. Tumor cells of ovarian papillary adenocarcinoma ^a [10]. Borderline, low-grade, and stage epithelial ovarian tumors, epithelial inclusion cysts ^b [103]. High-grade and stage serous ovarian cancer ^{a,b} [104]. Endometrial tumors ^a [105]		Stromal cells around a papillary adenocarcinoma ^a [10]	NR
Esophageal	Tumors and lymph node metastases ^{a,b} [106]		NR	NR
Head and neck	Primary tongue tumors ^a [107]. Head and neck squamous cell carcinomas ^b [54]		Primary and metastatic nasopharyngeal carcinoma, other head, and neck carcinomas ^{a,b} [78]	NR
Liver and pancreatic	Advanced stage, poorly differentiated hepatocellular carcinoma ^a [75]. Primary cholangiocarcinomas ^b [108]. Malignant pancreatic endocrine tumors with metastasis ^a [109]	Benign pancreatic endocrine tumors ^a [109]	Primary cholangiocarcinomas ^b [108]	NR
Prostate	Preneoplastic and malignant tumors ^b [110]		NR	NR
Thyroid	NR	NR	Aneuploid papillary thyroid cancer ^a [111]	NR
Mesenchymal tumors	Solitary fibrous tumors ^a [112]		NR	NR
Lymphoma and leukemia	Acute myeloid leukemia ^a [114, 115]. Primary CNS lymphoma ^a [176]	Non-CNS diffuse large B cell lymphoma ^a [176]	Primary Hodgkin's lymphoma ^{a,b} [116, 117]. AML ^a [114]	B cell non-Hodgkins lymphoma ^b [116]. Anaplastic large cell lymphoma ^{a,b} [117]

NR not reported

^aRNA^bProtein

6.1 Lung cancer

Among the human cancers, most of the advances in DDR research have been made in lung cancer, in particular in non-small cell lung carcinomas (NSCLCs). Indeed, several studies reported dysregulated expression, phosphorylation, and mutations of DDRs in NSCLCs. Moreover, recent evidence suggests that DDR2 represents a new therapeutic target in squamous cell carcinoma (SCC) of the lung [87]. Early studies by Ford et al. [88] in 146 primary NSCLC and in an independent set of 23 matched tumor and normal lung tissue samples showed that DDR1 mRNA was upregulated in tumor vs. normal tissue, whereas DDR2 was downregulated. Moreover, DDR1 was found to be an independent favorable prognostic marker for early-stage NSCLC patients while no association between DDR2 mRNA expression and prognosis or patient survival was observed [88]. In contrast, an immunohistochemical study using 171 NSCLC samples showed that DDR1 protein expression was associated with lymph node metastasis and poor overall survival [89]. Also, a recent study using a cohort of 83 patients with NSCLC found that tumors with high DDR1 protein levels (as determined by immunohistochemistry) were associated with poor survival [90]. Thus, two independent studies [89, 90] reported results that conflict with those of the Ford et al. study regarding the association of DDRs with lung cancer progression.

Dysregulated DDR function in NSCLC may be caused by altered receptor activation or the presence of mutations. Using a phosphoproteomic approach Rikova et al. [91] found that DDR1 and DDR2 are among the top 20 RTKs that are highly phosphorylated in a set of 150 NSCLC tumors. Specifically, DDR1 was the third most phosphorylated tyrosine kinase (following Met and Alk) while DDR2 was at position 13. Thus, enhanced DDR activation may be associated with development and/or progression of NSCLC. Several somatic *DDR1* and *DDR2* mutations have been identified in primary NSCLC samples. Davies et al. screened 26 primary NSCLC patient samples for somatic mutations in protein kinases and identified one somatic mutation in the *DDR1* gene, A496S, in one patient with squamous cell carcinoma and one somatic mutation in the *DDR2* gene, R105S, in one patient with large cell carcinoma [92]. These mutations map to the cytosolic JM region in DDR1 and the discoidin 1 domain in DDR2 (Fig. 1). No further validation or functional analyses of these mutations were performed. Ding et al. screened 188 primary lung adenocarcinoma samples for somatic mutations in 623 genes comprising known oncogenes, tumor suppressors, and protein kinase families and found two novel somatic mutations in the *DDR1* gene, W385C and F866Y, and one in *DDR2*, N456S, with each mutation occurring once in the cohort [93]. These mutations map to the extracellular JM and kinase domains in DDR1 and to the cytosolic JM domain in DDR2 (Fig. 1). A

recent study conducted to determine the mutational status of the entire tyrosine kinome of lung SCC using a set of 290 tumor samples and cell lines identified somatic mutations in the *DDR2* gene [87]. These analyses indicated a frequency of *DDR2* mutations at a rate of 3.8% in all samples (tumors and cell lines) and 3.2% in primary lung SCC samples. Overall, 11 mutations in *DDR2* were found throughout the entire gene and located in various DDR2 domains (Fig. 1). These include L63V, I120M, and D125Y within the collagen-binding discoidin 1 domain; L239R and G253C within the discoidin 2 domain; G505S in the cytosolic JM domain; and C580Y, I638F, T765P, G774E, and G774V within the kinase domain. Experimental data showed that the *DDR2* L239R and I638F mutations rendered SCC cell lines sensitive to the tyrosine kinase inhibitor dasatinib and conferred an oncogenic gain of function phenotype to NIH-3T3 fibroblasts in a colony-forming assay in soft agar. Wild-type DDR2 was also oncogenic in murine pro-B BA/F3 cells [87]. Interestingly, this study also reported that a SCC patient with no evidence of EGFR kinase mutations and who exhibited good therapeutic response to a combination of dasatinib and erlotinib harbored a *DDR2* mutation, S768R, which, based on modeling studies, could alter DDR2 kinase activity [87]. Based on these indirect findings, the authors suggested that the patient's response to the dasatinib/erlotinib regimen was due to a potential sensitivity to dasatinib caused by the mutant DDR2, since erlotinib was found to be a weaker inhibitor of DDR2. Together, these exciting results highlight the importance of further investigating the significance of dysregulated DDR function in the pathogenesis and treatment of NSCLC.

6.2 Breast cancer

Early studies in a few human breast cancer cell lines and in DDR1 knockout mice suggested that altered DDR1 expression and/or function is critical for breast cancer progression [1, 94]. However, conflicting results were reported when DDR1 mRNA expression was examined in breast cancer tissues. While one study reported abundant expression of DDR1 mRNA in primary invasive carcinoma and in lymph node metastases [95], another study showed a moderate reduction in DDR1 mRNA levels in the majority of intermediate to high-grade human breast carcinomas compared to normal mammary tissues [96]. These discrepancies may be due to differences in the histological type and/or tumor grade. Indeed, another study reported that DDR1 mRNA and protein levels were significantly decreased in invasive lobular carcinomas when compared to invasive ductal carcinomas [97]. While the functional significance of this observation for disease progression is still unknown, these intriguing results have potential clinical

application for distinguishing between invasive ductal and lobular carcinomas. DDR2 expression in human breast epithelial cells has been associated with induction of EMT [71], but the expression pattern and association of DDR2 with disease progression in breast cancer are unknown. Future studies will have to focus on determining DDR1 and DDR2 expression in the different subtypes of breast cancer and in the transition from *in situ* to invasive carcinoma.

6.3 Brain cancer

DDR1 mRNA was found to be highly expressed in high-grade pediatric [98] and adult brain tumors regardless of cell type (glioblastoma multiforme, anaplastic astrocytoma, anaplastic mixed glioma, primitive neuroectodermal tumors, ependymoma, and meningeal sarcoma) [99]. DDR1 protein expression was also found to be elevated in gliomas compared to normal brain tissue [100] and significantly correlated with poor clinical outcome [101]. Since the collagen matrix within the brain parenchyma is restricted to the vascular and perivascular areas, these findings suggest that in brain tumors DDR1 may play a role in invasion of glioma cells along the perivascular matrix. In pituitary adenomas, DDR1 protein is widely expressed, and its level correlates with the hormonal background of the tumor [102]. Specifically, DDR1 protein expression is significantly higher in growth hormone- and prolactin-producing adenomas compared to adrenocorticotropic-producing and non-functioning adenomas. DDR1 protein expression was also found to be higher in macroadenomas compared to microadenomas [102].

6.4 Gynecological cancers

An early study reported that DDR1 mRNA was expressed in moderately or poorly differentiated ovarian tumors, while benign or borderline tumors showed very little expression [8]. In a later study, high levels of DDR1 protein were found in all histological types of epithelial ovarian cancers and in ovarian epithelial inclusion cysts, which are thought to be sites of metaplastic changes within normal ovarian tissue. However, no significant correlation between DDR1 protein expression and prognosis was observed [103]. Another study in serous ovarian tumors found high levels of DDR1 protein in high-grade and advanced stage tumors compared to low-grade and early-stage tumors and a significant correlation between DDR1 protein expression and poor outcome [104]. Thus, two independent studies [103, 104] suggest that DDR1 may be a useful target and biomarker for early [103] and advanced [104] ovarian cancer. A study in endometrial cancer reported that DDR1 mRNA was significantly

upregulated in tumor tissues and in uterine aspirates, and suggested that DDR1 could serve as a novel diagnostic marker for this type of cancer [105].

6.5 Esophageal cancer

DDR1 mRNA and protein are highly expressed in esophageal carcinoma tissues [106]. In these tumors, DDR1 mRNA levels were found to significantly correlate with tumor cell proliferation. In addition, DDR1 protein was detected in infiltrating foci and lymph node metastases, suggesting a role for DDR1 in invasion and metastasis of this cancer type [106].

6.6 Head and neck cancers

DDR1 mRNA was found to be consistently upregulated in primary tongue tumors when compared to normal tongue mucosa [107]. Head and neck squamous carcinoma tissues were reported to express high levels of DDR1 protein [54]. Studies in nasopharyngeal carcinomas found high levels of DDR2 (mRNA and protein), but not DDR1, in the cancer cells of primary and metastatic tumors [78]. The high expression of DDR2 in these tumors was attributed in part to transcriptional activation of the *DDR2* promoter by the EBV-Z-transactivator protein [78].

6.7 Liver and pancreatic cancer

High levels of DDR1 mRNA were found in hepatocellular carcinomas, which significantly correlated with advanced tumor stage [75]. The increased expression of DDR1 mRNA in these tumors was associated with downregulation of miR-199-5p, a miRNA that directly targets the 3' UTR of DDR1 mRNA [75]. In cholangiocarcinoma tissues, both DDRs were identified in a phosphoproteomic screen for highly phosphorylated tyrosine kinases, with DDR1 being the third most phosphorylated tyrosine kinase in these tumors [108]. DDR1 was also identified as one of the 72 genes that were significantly upregulated in malignant versus benign pancreatic endocrine tumors [109].

6.8 Prostate cancer

DDR1 was found to be highly expressed in prostate cancer tissues and to significantly correlate with hormone independence, suggesting that it may play an important role in the development of the hormone refractory state in prostatic tumors [110]. Since advanced prostate cancer is characterized by development of bone metastasis, it will be also interesting to examine the functional contribution of DDRs to this process.

6.9 Thyroid cancer

Microarray analyses in aneuploid papillary thyroid carcinomas revealed that DDR2 is one of the few genes that are highly expressed in patients with metastatic disease at time of diagnosis [111]. Moreover, in the same study, analysis of tumors from patients that had died from this disease revealed that DDR2 was one of the most overexpressed genes in this group as well [111]. While these microarray results were validated by quantitative PCR, no further protein expression or functional analyses were performed.

6.10 Solitary fibrous tumors

Solitary fibrous tumors (SFT) are rare mesenchymal neoplasms that arise from soft tissues and are characterized by intense proliferation of spindle, fibroblast-like cells within a collagen stroma. A microarray study found upregulated DDR1 mRNA in SFT samples, regardless of anatomical location. However, sequencing of eight SFT samples showed no mutations in the *DDR1* gene [112]. Considering the role that DDRs play in regulating the interactions of mesenchymal cells with collagen, these results suggest that DDR1 may be involved in the development of SFT. Moreover, the high expression of DDR1 in a mesenchymal tumor disputes the notion that DDR1 and DDR2 should be classified as epithelial or mesenchymal receptors, respectively.

6.11 Lymphomas and leukemias

Dysregulated tyrosine kinase function is implicated in the pathogenesis of hematological malignancies. In fact, most of the targeted therapies in these cancer types are directed against kinases including RTKs. Therefore, a great effort has been invested in defining the signatures of kinases that are altered in hematological tumors. These searches identified the DDRs as one set of RTKs that are either overexpressed and/or mutated in hematological cancers. A study aimed at characterizing gene expression signatures in adult acute lymphocytic leukemia (ALL) reported that DDR1 was one of the highly expressed kinases in cases without evidence of molecular rearrangements and in ALL cases with BCR/ABL rearrangements. These studies suggested that dysregulated activation of DDR1 might play a role in the transformation of a subset of ALL [113]. Two studies reported the identification of DDR1 mutations in leukemias. Re-sequencing of 26 selected cytoplasmic tyrosine kinase and receptor tyrosine kinase genes in patients with *de novo* AML identified an A803V somatic mutation of *DDR1* in one out of the 188 patients tested [114]. This mutation maps to the predicted activation loop within the

DDR1 kinase domain suggesting the possibility that DDR1 activity is altered in AML [114]. In another study, Loriaux et al. [115] used high-throughput DNA sequencing to screen exons encoding the activation loop and cytosolic JM domains of 85 tyrosine kinase genes in 188 AML patients without *FLT3* or *c-KIT* mutations. These analyses identified five AML patients with non-synonymous somatic mutations in the DDR1 JM domain: T460M (1 sample), N502S (3 samples), and A533S (1 sample). However, these DDR1 mutations did not affect the kinetics or the extent of collagen-induced receptor phosphorylation when compared to wild-type DDR1 upon expression in BA/F3 cells. Also, these mutations failed to transform these cells into cells that exhibit hypersensitivity to interleukin-3-stimulated growth. However, due to the inherent limitations of the BA/F3 assay [115], the potential involvement of DDR1, alone, or in combination with other dysregulated RTKs, in AML cannot be ruled out and needs to be further explored.

A study in Hodgkin's lymphoma (HL) demonstrated enhanced expression of DDR2 protein in 30–70% of the Hodgkin/Reed–Sternberg (HRS) cells present in the lymph nodes of HL patients whereas the normal B cells showed no expression [116]. Interestingly, collagen I was detected near the HRS cells, suggesting that DDR2 can potentially be activated in these neoplastic cells. The high expression of DDR2 in HL was later confirmed in an immunohistochemical analysis of lymph node tissue infiltrated by HL, which showed high DDR2 in 44% of the HL samples [117]. Thus, DDR2 is emerging as one of the RTKs that are upregulated in HL. Collectively, these studies suggest that, as in solid tumors, dysregulated DDR expression or activity may contribute to the pathogenesis of hematological cancers, with DDR1 and DDR2 associated with leukemias and lymphomas, respectively.

7 Function of DDRs in normal and cancer cells and tissues

7.1 Functions of DDRs as revealed in knockout and mutant mice

The importance of DDRs in normal physiological functions has been demonstrated by the phenotypes of mice lacking either DDR1 or DDR2 [94, 118] and of mice carrying a spontaneous, autosomal-recessive mutation in the DDR2 gene [121]. All these mice exhibit major defects in skeletal development and consequently display dwarfism, suggesting that both DDRs play a critical role in bone development and growth. The role of DDR2 in skeletal development was further demonstrated with the identification of several mutations in the human DDR2 gene, which were shown to be the causative genetic defect of a human growth disorder characterized by disproportionate short stature and short limbs [119, 120]. The phenotype of DDR1- and DDR2-deficient

mice is also characterized by several abnormalities in the reproductive systems. For instance, DDR1 knockout mice exhibit impaired blastocyst implantation and aberrant mammary gland morphogenesis [94]. The mammary gland of DDR1 knockout mice displays delayed ductal elongation, abnormal secondary branching, hyperproliferative ductal epithelium, enlarged terminal end buds, and increased ECM deposition around ducts, and thus, these mice are unable to lactate [94], presumably due to the inability of the mammary epithelial cells to respond to lactogenic stimuli [124]. DDR2 mutant mice are infertile due to gonadal dysfunction including impaired spermatogenesis and ovulation, which were ascribed to defects in overall endocrine function [121–123]. Other organ systems are also altered in DDR1- and DDR2-deficient mice. For instance, DDR1 knockout mice exhibit altered kidney structure [125] and impaired primary mesangial cell adhesion and hyperproliferation due to increased mitogen-activated protein kinase/extracellular signal-regulated kinase (MAPK/ERK) activation [126]. These mice are also unable to control their ear movements [94] and show loss of auditory function with profound structural changes throughout the cochlear duct [127]. DDR2 knockout mice display impaired dermal wound healing due to defective proliferation, invasion, proteolytic activity, and ECM remodeling by skin fibroblasts [55, 128]. In addition, the DDR knockout mice exhibit altered responses to various insults and pathological conditions. For example, DDR1 knockout mice exhibit reduced neointimal thickening and altered VSM cell function following vascular injury. Also, these mice have reduced inflammatory and fibrotic responses in obstructive nephropathy, renal hypertension, bleomycin-induced lung injury, atherogenesis, and hereditary collagen IV disease (see Table 2 for details and references). Taken together, these findings suggest that DDR1 is involved in regulation of inflammatory and fibrotic responses in lung, vascular, and renal tissues. Similarly, DDR2 promotes HS cell activation and fibrosis in acute liver injury [79]. However, in chronic liver injury, DDR2 likely plays a suppressing role, since DDR2 knockout mice are more susceptible to inflammatory responses under this condition [129]. A recent report also found that DDR2 knockout mice are more prone to metastasis of colon carcinoma cells to the liver, presumably due to altered HS cell responses leading to a pro-metastatic environment within the liver tissue [130].

7.2 Functions of DDRs as revealed in cultured primary cells and cell lines

7.2.1 Cell proliferation and survival

Both DDR1 and DDR2 can exhibit pro- [30, 69, 79, 87, 101, 118, 128, 131–134] and anti- [94, 126, 129, 135, 136]

proliferative activities in a cell-type specific manner (summarized in Table 3). Additionally, DDR1 was found to promote survival of breast and colon cancer cell lines under conditions of genotoxic stress, thereby making these cancer cells resistant to chemotherapy [63, 64]. Studies in human HCT116 colon carcinoma cells showed that the pro-survival effects of DDR1 in response to genotoxic drugs are partly mediated by autocrine activation of Notch1 signaling in response to collagen-induced activation of DDR1 [52].

7.2.2 Differentiation and EMT

DDR1 have been shown to play key roles in regulation of cell differentiation in various cellular systems. DDR1 signaling is required for neuronal differentiation *in vitro* [137]. Also, DDR1 is one of the transcripts that are upregulated during neuronal differentiation of mouse embryonic stem cells [138]. DDR1 expression strongly correlates with oligodendroglial differentiation both *in vitro* and *in vivo* in a mouse model of remyelination [139]. Inhibition of DDR1 expression is associated with decreased osteogenic potential of human mesenchymal stem cells [85]. DDR1 promotes differentiation into myotubes of the skeletal muscle cell line, C2C12, in a process that requires kinase activity [65]. DDR1 expressing porcine LLC-PK1 kidney epithelial cells form microvilli even in the presence of collagen I (which promotes de-differentiation), while cells expressing a dominant negative DDR1 have impaired microvilli formation [59]. DDR1 expressing normal murine breast epithelial cells (NMuMG) and LLC-PK1 cells exhibit higher levels of E-cadherin and lower levels of the mesenchymal markers fibronectin, integrin $\beta 1$, and α -smooth muscle actin, suggesting a role for DDR1 in maintenance of the epithelial phenotype [59]. In contrast, in mouse embryonic stem cells, DDR1 signaling supports the de-differentiated state, possibly by cooperating with integrin $\alpha 2\beta 1$ signaling [66]. DDR2 promotes osteogenic differentiation in murine bone marrow stromal cells [61], pre-osteoblast MC3T3-E1 cells [61, 62], and pluripotent C3H10T1/2 cells and C3C12 cells [62]. DDR2 promotes chondrogenic differentiation in the murine chondrogenic cell line ATDC5 [61]. Transactivation of the transcription factor Runx2 by DDR2 is a key event in both osteogenic and chondrogenic differentiation in these systems [61, 62]. In adipocytes, however, DDR2 expression was found to inversely correlate with differentiation [140]. Thus, whether DDRs promote or inhibit differentiation appears to be cell type dependent. Regardless, given the roles that DDRs play in normal cell differentiation, it will be important to determine whether DDR signaling is involved in differentiation of cancer stem cells.

During normal development, certain epithelial cells acquire the ability to express mesenchymal properties in a

Table 2 Insights into DDR function from knockout mice and mutations in humans

	DDR1 knockout mice	DDR2 knockout mice	DDR2 mutations in mice (<i>slie</i>) and humans (<i>SMED-SL</i>)
Skeletal system	Smaller in size, poorly mineralized fibula bone [94]	Mouse: dwarfism and shorter long bones due to reduced chondrocyte proliferation [118]	Mouse: dwarfism [121] Human: multiple skeletal defects in <i>SMED-SL</i> [119], due to DDR2 trafficking defects or defects in collagen-binding activity [120]
Mammary gland	Lactational defect, delayed mammary gland ductal outgrowth, hyperproliferative epithelium, abnormal ductal branching, enlarged terminal end buds [94]. Reduced Stat5 activation and β -casein expression upon lactogenic stimulation in primary mammary epithelial cells [124]	NR	NR
Kidney	Thickening of glomerular basement membrane, focal loss of split diaphragms, proteinuria [125]. Reduced adhesion of primary mesangial cells to ECM, increased mesangial proliferation due to increased MAPK activity [126]	NR	NR
Vascular system	Reduced neointimal thickening after vascular injury [30]. Reduced adhesion and migration to collagen I and MMP secretion by SM cells [147]. Reduced atherosclerotic calcifications in <i>Ldlr</i> ^{-/-} background [177]. Increased matrix secretion, proliferation, and migration of vessel wall SM cells [135]	No effect on adhesion, migration, or proliferation of vascular SM cells [152]	NR
Inflammatory/fibrotic response/immune system function	Reduced inflammatory response to LPS, reduced glomerular fibrosis, and inflammation in hypertension [178]. Impaired macrophage migration and invasion in response to MCP-1 [150, 157]. Reduced inflammatory and fibrotic response to bleomycin-induced lung injury, no p-38 MAPK activation [179]. Reduced macrophage recruitment, inflammation, and fibrosis in atherogenesis in <i>Ldlr</i> ^{-/-} background [180]. Reduced proinflammatory and profibrotic cells in kidney in hereditary collagen IV disease (<i>COL4A3</i> ^{-/-} background) [181]. Reduced inflammation and fibrosis in obstructed kidneys [157]	Increased inflammatory and proliferative response of hepatic stellate cells and macrophages to chronic liver injury [129]	NR
Skin	NR	Reduced proliferation, invasion through basement membrane, and MMP activity of dermal fibroblasts [128]. Impaired dermal wound healing due to defective ECM remodeling by skin fibroblasts [55]	NR
Ear	Unable to control ear movement [94]. Loss of auditory function and profound structural changes throughout the cochlear duct [127]	NR	NR
Reproduction	Impaired blastocyst implantation into uterine wall [94]	NR	Mouse: impaired spermatogenesis [121, 123] and ovulation [121, 122] in <i>slie</i> mice
Susceptibility/resistance to diseases	Resistant to bleomycin-induced lung fibrosis [179]. Resistant to hypertension-induced renal failure [178]. Decreased atherosclerosis in <i>Ldlr</i> ^{-/-} background [177, 180]. Delayed renal fibrosis, inflammation, and death in hereditary collagen IV disease (<i>COL4A3</i> background) [181]	Increased susceptibility of hepatic tissue to colon carcinoma metastasis [130]. Increased susceptibility to hepatic fibrosis in chronic liver injury [129]	NR

NR not reported, *SMED-SL* short-limb abnormal calcification, *SM* smooth muscle

Table 3 DDR functions reported in cultured cells

	DDR1		DDR2	
	Promoting	Suppressing	Promoting	Suppressing
Survival and Chemoresistance	Non-malignant cells: MDCK cells [56] Malignant cells: human prostate cancer cells (PC3, DU145, LNCaP) [110], human breast cancer cells (MCF7 [63], T47D [64]), human colon cancer cells (HCT116) [52, 63]	Non-malignant cells: primary murine mesangial cells [126], murine VSM cells [135]	NR	NR
Proliferation and colony-forming ability	Non-malignant cells: murine VSM cells [30], primary HBECs [131], T cells [132] Malignant cells: human glioma cells (U251, G1-1, T98G) [101], HCT116 (<i>in vitro</i> colony-forming assay) [52]	Non-malignant cells: rat HS cells [79], murine skin fibroblasts [118, 128], rat VSM cells [133], murine chondrocytes [118], Slug-MDCK cells [69], NIH-3T3 fibroblasts can be oncogenically transformed by DDR2 L63V and I638F mutations [87] Malignant cells: human melanoma cells (A375), hepatoma cells (SK-HEP), colon carcinoma cells (HT-29) [134], human lung squamous cell carcinoma (H12286, HCC366) [87]	Non-malignant cells: rat HS cells [79], murine skin fibroblasts [118, 128], rat VSM cells [133], murine chondrocytes [118], Slug-MDCK cells [69], NIH-3T3 fibroblasts can be oncogenically transformed by DDR2 L63V and I638F mutations [87] Malignant cells: human melanoma cells (A375), hepatoma cells (SK-HEP), colon carcinoma cells (HT-29) [134], human lung squamous cell carcinoma (H12286, HCC366) [87]	Non-malignant cells: murine HS cells after chronic liver injury [129], 3T3-L1 pre-adipocytes [140] Malignant cells: fibrillar collagen-induced proliferation in human melanoma cells (M24met, A2058), human fibrosarcoma cells (HT1080) [136]
Differentiation	Non-malignant cells: murine granule neuron axon formation [137], osteogenic potential in human mesenchymal stem cells [85], epithelial differentiation in porcine kidney epithelial cells (LLC-PK1) and murine mammary epithelial cells (NMuMG) [59], murine myoblast (C2C12), differentiation/formation of myofibers [65]	Non-malignant cells: maintenance of undifferentiated state in murine embryonic stem cells [66]	Non-malignant cells: differentiation in murine osteoblasts [61, 62], and chondrocytes [61]	Non-malignant cells: differentiation of murine pre-adipocytes to functional adipocytes [140]
EMT	Malignant cells: Collagen I-induced cell spreading in human pancreatic cancer cells (BxPC3) [48], TGF β /collagen I-induced EMT in human lung carcinoma cells (A549) [144]	Non-malignant cells: DDR1 expression suppresses mesenchymal genes in LLC-PK1 and NMuMG [59]	Non-malignant cells: TGF β /collagen I-induced EMT in renal proximal tubule cells (HK-2) [144] Malignant cells: TGF β /collagen I-induced EMT in A549 [144]	Non-malignant cells: differentiation of murine pre-adipocytes to functional adipocytes [140]
Cell-cell contact	Non-malignant cells: MDCK [53, 57], LLC-PK1, NMuMG [59] Malignant cells: human oral squamous carcinoma cells (A431, SCC12) [54]		NR	NR
Adhesion	Non-malignant cells: murine VSM cells [30, 147], human mammary epithelial cells (HB2) [38, 39], human monocytic cells (THP-1) [148], primary murine mesangial cells [126], primary human melanocytes [149], murine macrophages [150] Malignant cells: human pituitary adenoma cells (HP-75) [102], human glioma cells (G140) [100], human hepatoma cells (Huh7) [151]		NR	NR
Spreading		Non-malignant cells: NIH3T3 fibroblasts [47], human mesenchymal stem cells [85], MDCK [57, 58] Malignant cells: MCF7 [47]	NR	NR
Migration	Non-malignant cells: murine VSM cells [30, 41, 147], THP-1 cells [148], human T cells [67, 156], NIH3T3 fibroblasts, mouse	Non-malignant cells: HB2 cells [38], MDCK [45, 56], LLC-	Non-malignant cells: murine skin fibroblasts [128], rat VSM cells [76, 77, 133]	Non-malignant cells: murine HS cells and macrophages

Table 3 (continued)

	DDR1	DDR2	
	Promoting	Suppressing	
	Promoting	Suppressing	
	embryonic fibroblasts [47], murine macrophages [157], HBECs and human bronchial epithelial cells (BEAS-2B) [131] Malignant cells: G140 cells [100], human hepatocellular carcinoma cells (HLE, Huh 7) [158], human lung adenocarcinoma cells (A549, H538) [89], human lung large cell carcinoma cells (H460) [90], human breast carcinoma cells (MDA-MB-231) [159] (T47D, MDA-MB-468) [96], (MCF7) [47]	PK1 [59], murine VSM cells [135] Malignant cells: Huh7 cells [151], MCF7 (when DARPP32 is expressed), MDA-MB-231 (when DDR1 and DARPP32 are expressed) [49]	from chronically injured liver [129]
Invasion	Non-malignant cells: murine macrophages [150] Malignant cells: HP-75 cells [102], A431 and SCC12 cells [54], G140 cells [100], U251, GI-1, and T98G cells [101], HLE and Huh 7 cells [158], human hepatoma cells (HEPG2, SNU-182) [75], PC3, DU145 and LNCaP cells [110], A549 and H538 cells [89], H460 cells [90], MDA-MB-231 cells [161]	Non-malignant cells: rat HS cells [79]	
Collagen remodeling	Non-malignant cells: organization of native collagen I matrix by human mesenchymal stem cells [85]	Non-malignant cells: human VSM cells (collagen I degradation) [182], murine skin fibroblasts (collagen I deposition) [55, 128]	Non-malignant cells: soluble DDR2 extracellular domains suppress collagen I fibrillogenesis [84, 173, 183]
MMP expression	Non-malignant cells: murine VSM cells (MMP2, 9) [30, 147], human VSM cells (MMP1) [182], LDLR ^{-/-} murine macrophages (MMP2, 9, 14) [180], HBECs and BEAS-2B cells (MMP7) [131]	Non-malignant cells: rat HS cells (MMP2) [79], murine skin fibroblasts (MMP2 secretion) [55, 128], rat VSM cells (MMP2) [77, 133], human VSM cells (MMP1, MMP2) [182], osteoarthritic chondrocytes in collagen XI ^{-/-} mice (MMP13) [184], murine chondrocyte cell line C-28/12 (MMP13) [185], human rheumatoid arthritis synovial fibroblasts (MMP1) [186], (MMP13) [187], NIH-3T3 cells (MMP-1) [186], murine synovioocytes and 293T cells (MMP13) [187], primary goat articular chondrocytes (MMP13) [60] Malignant cells: A375 cells (MMP2, 9) [134]	Non-malignant cells: increased metastasis to liver of DDR2 knockout mice [130]
Tumor growth and metastasis	Malignant cells: HP-75 cells (MMP2, 9) [102], G140 cells (MMP2) [100], HLE and Huh 7 cells (MMP2, 9) [158], PC3, DU145, and LNCaP cells (MMP9) [110], H538 cells (MMP9) [89], MDA-MB-231 cells (MMP-2, 9) [161] Malignant cells: HCT116 (xenografts) [52], H460 metastasis to bone [90]	Malignant cells: A375 metastasis to liver [134]	

NR not reported, VSM vascular smooth muscle, HBECs human bronchial epithelial cells, HS hepatic stellate

phenomenon known as EMT. EMT occurs during such processes as gastrulation, neural crest formation, pancreas, liver and reproductive tract development, atrioventricular canal formation in the heart, and during physiological responses to injury, just to name a few (reviewed in [141]). EMT is characterized by the activity of specific transcription factors, which, among other effects, induce a switch in cadherin expression (from E-cadherin in the epithelial state to N-cadherin in the mesenchymal state) and a loss of cell polarity. EMT has been associated with increased cell motility and invasiveness, induction and maintenance of stem cell properties, prevention of apoptosis and senescence, resistance to immuno/chemotherapy, and escape from immune surveillance in pathological conditions such as fibrosis and tumor progression (reviewed in [141]). Induction of an EMT in various cellular systems has been found to correlate with differential DDR expression, from DDR1 (epithelial) to DDR2 (mesenchymal). For instance, EMT during heart development in chicken embryos is associated with high levels of DDR2 mRNA and protein [142]. In mice, elevated DDR2 mRNA was found in epicardium-derived cells undergoing EMT during development of the annulus fibrosis of the heart [143]. EMT induction in MDCK cells by Slug causes a switch from DDR1 to DDR2, which is thought to be responsible for the increased proliferation of these cells within collagen I gels [69]. TGF- β -induced EMT in primary human bronchial epithelial cells reduces DDR1 protein levels while DDR2 levels are unchanged [70]. Likewise, TGF- β induces DDR2 expression in renal proximal tubule epithelial cells, and knocking down DDR2 expression suppresses TGF- β or collagen I-induced EMT in these cells [144]. Expression of EMT inducers (Goosecoid, Snail, Twist, Slug, and TGF- β) in a non-transformed human mammary epithelial cell line induces an “EMT core signature,” which includes upregulation and downregulation of DDR2 and DDR1, respectively [71]. Downregulation of DDR1 was attributed to the presence of Zeb1 binding sites within the *DDR1* promoter. Zeb1 is a transcription factor that plays a critical role in mediating gene expression changes during EMT, particularly in E-cadherin downregulation [145]. Gene expression analyses in human breast cancer cell lines showed that DDR1 mRNA is lower in the more mesenchymal and invasive basal B type cell lines when compared to the basal A and luminal breast cell cancer lines [146]. While these studies suggest that a mesenchymal-like phenotype correlates with downregulation of DDR1, other studies suggest that, both DDRs are associated with EMT. For instance, knockdown of either DDR1 or DDR2 inhibits TGF- β - or collagen I-induced EMT in human A549 lung carcinoma cells [144] while in a pancreatic cancer cell line, Bx-PC3, DDR1 cooperates with integrins to mediate collagen I-induced EMT [48]. Thus, the relationship between induction or maintenance

of EMT and DDRs cannot be generalized and may be context-dependent. Regardless, DDRs may contribute to the adaptation of cells to the collagen environment during EMT-induced migration programs.

7.2.3 Cell adhesion and spreading

Regulation of cell adhesion is a major biological function of DDRs. Indeed, a plethora of studies demonstrated that both DDR1a and DDR1b support adhesion to collagen I in a variety of cell types [30, 39, 126, 147, 148]. Moreover, DDR1-mediated cell adhesion to collagen appears to be essential for tissue and cellular functions in normal and pathological processes. For instance, in melanocytes, DDR1-mediated adhesion to collagen IV plays a key role in the localization of these cells to the BM/basal layer of the epidermis [149]. In macrophages, DDR1 supports monocyte chemotactic protein (MCP)-1 induced adhesion to and invasion through collagen IV, which may contribute to macrophage infiltration and accumulation during atherogenesis [150]. DDR1 has been shown to promote adhesion to collagen of a variety of malignant cells including pituitary adenoma [102], glioma [100], and hepatoma cells [151]. In contrast, a recent study showed that DDR1 had no effect on the adhesion of human lung large cell carcinoma cells to collagen I or IV [90]. In the case of DDR2, its contribution to normal or cancer cell adhesion is still unclear. For example, one study showed that DDR2 is not required for adhesion of VSM cells to collagen I [152], while another study showed that DDR2 expression and activation plays an important role in regulating FAK levels in VSM cells attaching to collagen I [153].

The role of DDR phosphorylation (and the subsequent activation of downstream effectors) in cell adhesion has not been studied in detail and thus is barely known. A study using murine VSM cells showed that adhesion to collagen I is effectively mediated by kinase inactive DDR1, suggesting that in this system, kinase activity of the receptor is not critical [147]. However, whether this is the case for other cell types and collagens remains to be determined. Regardless, while molecular interactions between DDRs and collagen are not dependent on receptor phosphorylation [21–23], it is reasonable to postulate that meaningful cellular adhesion to collagen (as defined by the establishment of cell–matrix contacts, changes in cell morphology, and activation of downstream effectors) requires receptor structural changes and phosphorylation of specific tyrosine residues.

The interaction of adhesion receptors with the ECM initiates a cascade of signaling events, which subsequently triggers actin polymerization and myosin contraction, resulting in active cell spreading [154, 155]. To date, all reports show that DDR1 activity suppresses cell spreading in a variety of cell types [47, 57, 58, 85]. For instance, expression of

DDR1 was shown to inhibit spreading of MDCK cells plated on collagen [57] via inhibition of $\alpha 2\beta 1$ integrin-mediated Cdc42 activity [58]. In NIH3T3 fibroblasts and MCF7 breast cancer cells, DDR1 was shown to inhibit cell spreading via interaction with NMHC-IIA, a contractile protein involved in cell spreading [47]. At present, the role of DDR2 in cell spreading is unknown.

7.2.4 Cell–cell adhesion

Evidence suggests that DDR1 plays a role in supporting cell–cell interactions in epithelial cells independently of its ability to bind collagen. DDR1 was found to localize to cell–cell junctions, usually with E-cadherin, in a variety of normal [53, 54, 57, 59] and malignant [54] cell lines and tissues. Both DDR1 and DDR2 were shown to interact with E-cadherin via their extracellular domains [53]. In the case of DDR1, a complex with E-cadherin abrogates collagen-induced DDR1 activation, possibly by sequestering the receptor away from its ligand [57]. Conversely, DDR1 expression stabilizes E-cadherin at the cell surface and promotes cell–cell aggregation [53] by negatively regulating Cdc42 [59]. DDR1 also promotes cell–cell adhesion by reducing actomyosin contractility at cell–cell junctions, independent of receptor activation since neither the kinase inactive (K618A) nor the collagen-binding (R105A) DDR1 mutants has any effect on this function [54]. Following recruitment to cell–cell junctions by E-cadherin, DDR1 associates with the Par3/Par6 proteins via its C-terminal, PDZ interacting domains. The Par3/Par6 complex in turn, through RhoE recruitment, abrogates RhoA/ROCK activity and actomyosin levels at the cell–cell junction. This function of DDR1 was found to be indispensable for collective invasion by human oral squamous carcinoma cells [54]. In the case of DDR2, the functional significance of the E-cadherin/DDR2 complex [53] remains unknown.

7.2.5 Cell migration

DDRs regulate cell migration in a cell-type and isoform-dependent manner. For instance, in activated human T cells, DDR1 supports amoeboid migration within 3D collagen I lattices [67, 156]. Likewise, overexpression of DDR1a, but not DDR1b, enhances migration of THP-1 monocytic cells through 3D collagen I [148]. DDR1 is also required for the migration of primary human bronchial epithelial cells and the human bronchial epithelial cell line BEAS-2B [131]. In addition, DDR1 expression increases the migration of mouse embryonic fibroblasts and NIH3T3 fibroblasts toward collagen I [47]. Consistent with a pro-migratory activity of DDR1, macrophages isolated from DDR1 knockout mice exhibit impaired chemotactic migration in response to MCP-1 [157], and VSM cells derived from these mice exhibit impaired migration toward collagen I, which is

restored by expression of DDR1b [30, 41, 147]. DDR2 also supports the migration of rat VSM cells under hypoxic conditions via the p38 MAPK pathway [76, 77], and re-expression of DDR2 in DDR2-deficient murine fibroblasts restores cell migration through Matrigel [128]. Promotion of cell migration by DDR1 was also reported in various cancer cell lines including glioma [100], hepatocarcinoma [158], lung [89], and breast [47, 96, 159]. In some breast cancer cell lines (T47D and MDA-MB-468), DDR1-dependent promotion of cell migration was ascribed to regulation of the migration suppressor Syk kinase [96]. DDR2 was shown to support the migration of human A375 melanoma cells, SK-HEP hepatoma cells, and HT-29 colon carcinoma cells through collagen I [134].

Other studies showed that DDrs suppress cell migration. Overexpression of DDR1a or DDR1b in MDCK cells markedly reduces collagen I-induced migration in a process that involves inhibition of Stat1 and Stat3 tyrosine phosphorylation [45, 56]. Knockdown of DDR1 in MDCK cells or expression of dominant negative DDR1 in the porcine kidney epithelial cells, LLC-PK1, dramatically increases migration toward collagen I [59]. In the hepatoma cell line, Huh7, DDR1a overexpression reduces migration toward collagen I, fibronectin, and Matrigel [151]. In non-malignant human mammary epithelial cells, Wnt-5a inhibits migration possibly through activation of DDR1 [38] while in the breast cancer cell lines MCF7 and MDA-MB-231, DDR1 apparently suppresses migration only when co-expressed with its interacting partner, the phosphoprotein DARPP32 [49]. Interestingly, both Wnt-5a and DARPP32 are highly expressed in non-malignant mammary epithelial cells, but their expression is reduced or absent in breast cancer cell lines such as MCF7 and MDA-MB-231 [38, 49]. These observations suggest that the effect of DDR1 on cell migration, at least in mammary epithelial cell lines, is dependent on expression of co-factors such as Wnt-5a and DARPP32.

7.2.6 Cell invasion

The ability to invade through ECM and, consequently, to disseminate to other organs is a characteristic of malignant cancer cells. Invasion of ECM is a complex process requiring the activation of multiple genes but eventually depends on the action of key molecules such as ECM receptors and ECM-degrading proteases. Most studies investigating the role of DDrs in invasion utilize Matrigel, a tumor-derived basement membrane extract. Although Matrigel is rich in laminin and collagen IV, two major components of basement membranes, its relevance for the study of cell invasion has been questioned, particularly due to the lack of a covalently cross-linked collagen IV network, as is the case in natural basement membranes [160]. Therefore, more studies are required to determine the relative contribution of DDrs

in models of cell invasion that better mimic the structure and composition of natural ECMs, which may influence DDR signaling. Nevertheless, accumulating evidence using Matrigel as matrix barriers suggests that DDR1 plays a promoting role in invasion of a variety of human cancer cell lines including prostate [110], breast [161], lung [89], pituitary adenoma [102], oral squamous cell carcinoma [54], hepatocellular carcinoma [75, 158], and glioma [100, 101]. The pro-invasive activity of DDR1 in the Matrigel assay appears to be mediated by upregulation of MMP expression, in particular MMP-2 and MMP-9, which may contribute to degradation of matrix components [89, 100–102, 110, 158, 161]. Thus, DDR1 cooperates with the proteolytic machinery of the cells to facilitate invasiveness. However, how DDR1 regulates MMP expression and activity is unknown. Comparison of the respective roles of DDR1a and DDR1b showed that while in glioma cell lines DDR1a but not DDR1b is necessary for cellular invasion [100], both isoforms support the invasion of lung [89] and hepatocellular carcinoma cell lines [158]. The differential effects of DDR1 isoforms on invasion may be due to the activation of distinct signaling networks, which consequently may control invasive activity in a cell-type specific manner. However, the downstream effectors involved in regulating DDR1 function in tumor cell invasion have not been studied in detail and remain to be elucidated. A recent study suggested that the ability of DDR1 to support collective cell invasion of human A431 oral squamous cell carcinoma cells does not require receptor signaling and is independent of collagen binding. In these cells, DDR1 supports collective cell migration by its ability to regulate actomyosin contractility at sites of cell–cell junctions via formation of a complex with E-cadherin. Consequently, downregulation of DDR1 leads to a loss of cell–cell adhesion and decreased collective cell invasion [54]. In this mode of invasive behavior, DDR1 may contribute to cell invasion by acting as a scaffolding protein to sustain cell–cell interactions, independent of its ability to signal in response to collagen. Thus, it is expected that the impact of DDR1 on the invasive phenotype of cancer cells will be rather complex involving both collagen-dependent and collagen-independent mechanisms.

7.2.7 Tumor growth and metastasis

Up to this writing, only three studies have addressed the ability of DDRs to promote tumor growth and metastasis in animal models. Downregulation of DDR1 expression in HCT116 human colon cancer cells reduced size and weight of subcutaneous tumors in athymic mice [52]. Similarly, downregulation of DDR1 in human H460 lung large cell carcinoma cells substantially reduced tumor metastasis to bone and significantly decreased bone tumor burden after intratibial injection [90]. DDR1 appears to promote homing

of tumor cells to the bone, survival within the osseous environment and to increase osteolytic lesions, possibly by promoting osteoclastogenic activity [90]. However, how DDR1 regulates tumor-derived osteoclastogenesis remains unclear. Downregulation of DDR2 in A375 human melanoma cells reduced the amount of tumor burden in livers after intrasplenic inoculation of the cells [134]. Interestingly, while tumor-derived DDR2 appears to support the ability of melanoma to metastasize to the liver [134], host (hepatic) derived DDR2 appears to elicit a metastatic suppressive phenotype [130]. This anti-metastatic effect of host DDR2 was ascribed to an enhanced immune suppressive and profibrogenic response in the liver of DDR2 knockout mice resulting in the generation of a pro-metastatic niche and increased liver metastasis of MCA38 mouse colon cancer cells [130]. While the significance of these studies in cancer patients needs to be established, these findings highlight the potential contribution of stromal derived DDRs to cancer progression.

A few studies in human cancer tissues examined the association between DDR expression and disease progression. Yang et al. found that DDR1 expression in 171 samples of NSCLCs significantly correlated with lymph node metastasis and with reduced overall survival [89]. Similarly, DDR2 expression in aneuploid papillary thyroid carcinomas was associated with presence of distant metastasis and death from disease [111]. It is evident that more studies are required to establish the association and relative contribution of DDRs to cancer progression both in human cancers and in experimental models of cancer. However, such studies will have to make a distinction between DDR1 isoforms and DDRs with specific mutations, which may generate different outcomes in different tumor types. Moreover, the use of transgenic and DDR knockout mice with altered expression of known oncogenes and tumor suppressors in different tissues will provide a glimpse into the relationship and cross talk between cancer-initiating genes and DDR signaling at various stages of cancer progression.

8 DDRs as therapeutic targets in cancer

Given the accumulating evidence implicating DDRs in cancer progression, these receptors may represent promising new targets for specific tyrosine kinase inhibitors (TKIs). Approaches that target tyrosine kinase activity and function involve small chemical molecules designed to disrupt intracellular kinase activity or blocking monoclonal antibodies designed to interfere with ectodomain function. Small molecule TKIs are mostly ATP-competitive inhibitors, which bind to either the active (type I inhibitors) or inactive (type II inhibitors) conformation of the kinase thereby interfering with the transfer of the terminal phosphate of ATP to proteins containing a tyrosine residue [162, 163]. The

conserved structure of the kinase domain represents a significant challenge for the design of small molecule TKIs, and thus, many of the TKIs used today in the clinic are known to target multiple kinases including DDRs [164–166]. Chemical proteomic profiling studies identified imatinib [165], nilotinib [164], and dasatinib [164], as DDR1 inhibitors (Table 4). These TKIs inhibit collagen-induced activation of DDR1b and DDR2 expressed in HEK293 cells with IC_{50} values in the low nanomolar range [167]. Immobilized dasatinib has also been shown to preferentially capture DDR1 from lung cancer tissues and cell lines [168]. Moreover, dasatinib was shown to inhibit mutated DDR2 in lung cancer cells both *in vitro* and *in vivo* [87], suggesting that lung cancer patients harboring mutated DDR2 may benefit from dasatinib therapy. Another small TKI of DDRs is bafetinib (INNO-406), an inhibitor of the BCR–ABL, PDGFR, KIT, and LYN kinases and a structural analog of imatinib and nilotinib, [169]. LCB 03-0110, a thienopyridine derivative, was identified from a chemical library using the kinase domain of DDR2 and has been shown to inhibit collagen-induced DDR1 and DDR2 activation in HEK293 cells. However, this compound is also an effective inhibitor for other RTKs [170]. Together, these studies show that DDRs are high-affinity targets of many established and new small molecule TKIs, and it is therefore reasonable to assume that some of the anti-tumor activities, as well as some of the side effects [171], of small molecule TKIs currently used in cancer patients are due in part to interference with DDR signaling. However, this will not be the case in tumors that are not driven by DDR activity. For instance, mutation of the “gatekeeper” residue [163] in the kinase domain of DDR1 did not confer resistance to dasatinib in glioma [172] and lung cancer [168] cells, which are driven by members of the Src family of kinases (which are high-affinity targets of dasatinib). Nonetheless, these dasatinib-resistant DDR “gatekeeper” mutants [168, 172] represent new and valuable tools to evaluate the relative role of DDRs in various cancers. Assessment of the relative contribution of DDRs to cancer progression will require development of highly selective DDR kinase inhibitors. This effort will require elucidation of the unique structural features of the kinase domain of DDRs and an understanding of their structural/

functional dynamics in a cellular context that includes collagen as the ligand. In addition, kinase inhibitors for DDRs will have to be examined in tumors harboring defined DDR mutations [87, 92, 93, 114, 115].

Inhibition of kinase activity is not the only option to block DDR function in cancer. The soluble ectodomains of DDRs have been shown to interfere with endogenous receptor function [156] and to exhibit collagen binding and biological functions [17, 22, 25, 84, 173]. Thus, targeting the ectodomain of DDRs is a viable approach to neutralize receptor function in cancer cells. A search of chemical compounds able to inhibit the interactions of collagen type I with soluble DDR2 expressed in insect cells led to the identification of actinomycin D as an inhibitor of DDR2 [174]. Actinomycin D blocked collagen-induced DDR2 activation in HEK293 cells but had no effect on DDR1 activation. The inhibitory mechanism of DDR2 activation by actinomycin D remains unclear. Although a rational effort to produce function-blocking monoclonal antibodies against DDRs has not been reported, two commercially available antibodies from Santa Cruz Biotechnology were reported to block either DDR1a-mediated invasion of Matrigel and collagen adhesion of glioma cells [100] or receptor activation in human MDA-MB-231 cells [159] (Table 4).

9 Conclusions

Recent discoveries in DDR actions and the observation that DDRs can be mutated in cancers have raised considerable interest in these receptors, particularly as potential therapeutic targets. Although new and exciting findings in the DDR field are now beginning to emerge, many questions remain unanswered: (1) What is the profile of DDR expression in various cancer types? (2) Does DDR expression and/or mutational status correlate with disease progression and survival? (3) Can we utilize DDR mutational studies to predict patient outcome and determine treatment? (4) What is the contribution of DDRs (wild type and mutated) to tumor growth and metastasis in preclinical models of cancer? (5) What is the relative functional contribution of

Table 4 Reported DDR inhibitors

Inhibitor	Type	Target
Dasatinib [164]	Kinase type I inhibitor	DDR1 and DDR2
Nilotinib [164]	Kinase type II inhibitor	DDR1 and DDR2
Imatinib (Gleevec®) [165]	Kinase type II inhibitor	DDR1 and DDR2
Bafetinib (INNO-406) [169]	Kinase type II inhibitor	DDR1 and DDR2
LCB 03-0110 [170]	Thienopyridine derivative	DDR1 and DDR2
Actinomycin D [174]	Antibiotic	DDR2
Antibodies 48B3 [100] and H-126 [159]	Blocking antibodies	DDR1

DDR1 isoforms in cancer progression? (6) What underlies the unusual kinetics of DDR phosphorylation and what are the signaling pathways activated by wild-type and mutated DDRs in response to the different collagen microenvironments? (7) What is the profile of the protein interactome that regulates DDR activity in cancer cells? (8) What are the structural features of DDRs that define their downstream signaling effectors? (9) What are the posttranslational mechanisms (such as trafficking, endocytosis, and proteolysis) that regulate DDR activity in cancer cells? (10) Is there a DDR switch during EMT? (11) Are DDRs viable therapeutic targets in cancer? And if so, what kind of DDR specific inhibitors can we develop? The answers to these questions will shed light into the action of a unique set of RTKs that have the promise to become novel therapeutic targets in cancer.

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Conflict of interest The authors declare that they have no conflict of interest.

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